



# Prostate tumorigenesis in estrogen receptor $\beta$ -inactivated, prostate targeted fibroblast growth factor 8b-transgenic mice

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## Introduction

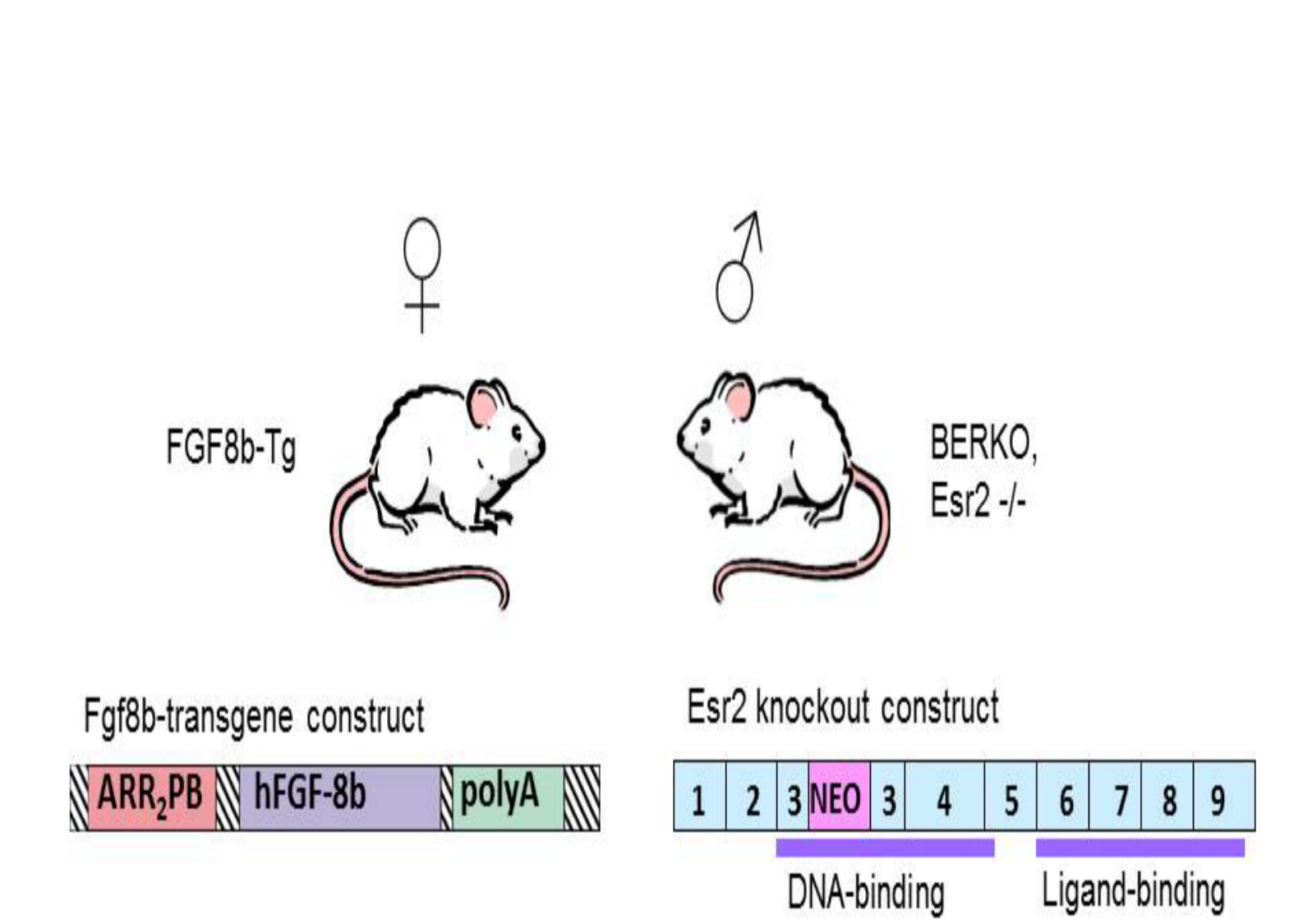
In the prostate, *Esr1* has been suggested to mediate tumor-promoting and *Esr2* anti-tumorigenic functions. *Esr2* knockout (BERKO) mice have been reported to generate prostate hyperplasia as well as increased proliferation, inflammation and decreased differentiation of epithelial cells in the prostate. Fibroblast growth factor 8 (FGF-8) is a mitogenic, angiogenic and transforming growth factor, that has four isoforms in human (a, b, e, f). The level of FGF8 has been found to be elevated in breast, ovarian and prostate cancer as well as in premalignant prostatic intraepithelial neoplasia (PIN) lesions. *Fgf8b*-transgenic (*Fgf8b*-Tg) mice develop advancing stromal and epithelial prostatic changes that slowly progress to mouse PIN (mPIN) lesions and to prostate cancer with mixed features of adenocarcinoma and sarcoma at old age (Elo et al. 2010).

Our objective was to study whether inactivation of *Esr2* affects prostate tumorigenesis, inflammation and stromal changes observed in *Fgf8b*-Tg mice.

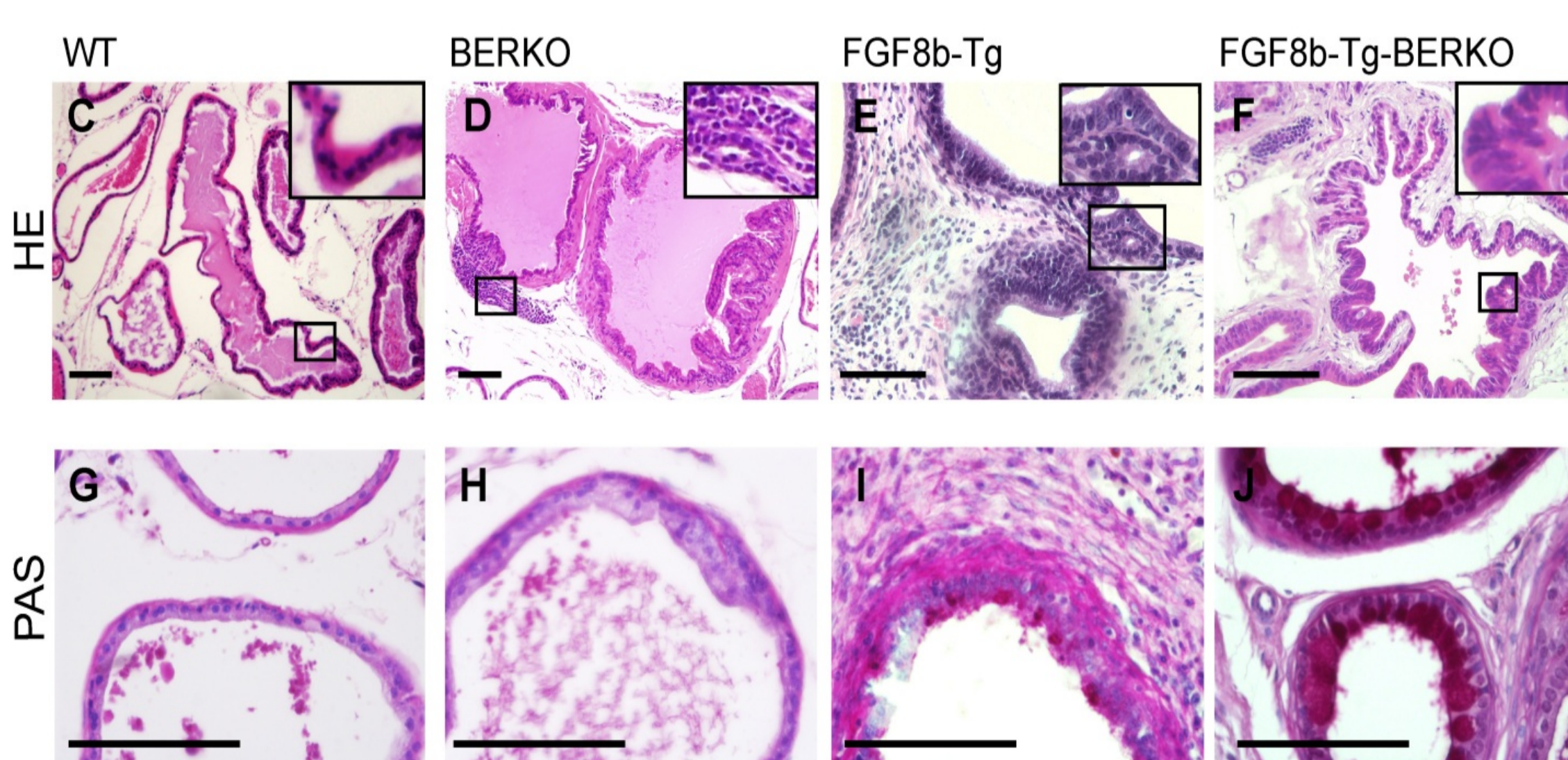
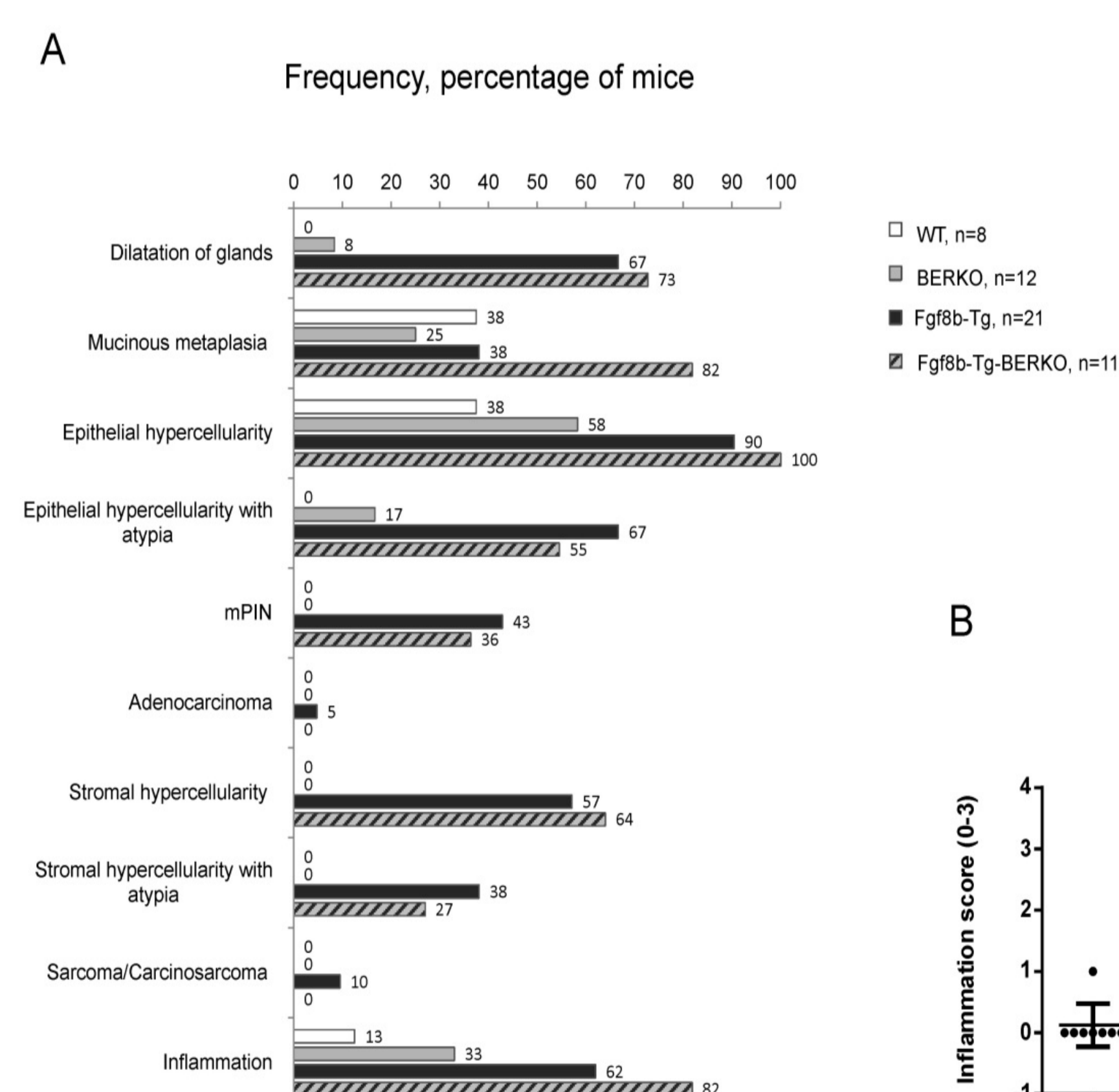
## Materials and Methods

BERKO mice (Krege et al. 1998) were bred with prostate targeted *Fgf8b*-Tg mice previously generated by us, to obtain *Fgf8b*-Tg-BERKO mice bearing two genomic modifications (Fig 1). Prostate histology of over 12-month-old WT, *Fgf8b*-Tg, BERKO and *Fgf8b*-Tg-BERKO mice were analyzed. Quantitative RT-PCR (qRT-PCR) and immunohistochemical (IHC) stainings are used to study gene expression and presence of proteins in the prostate.

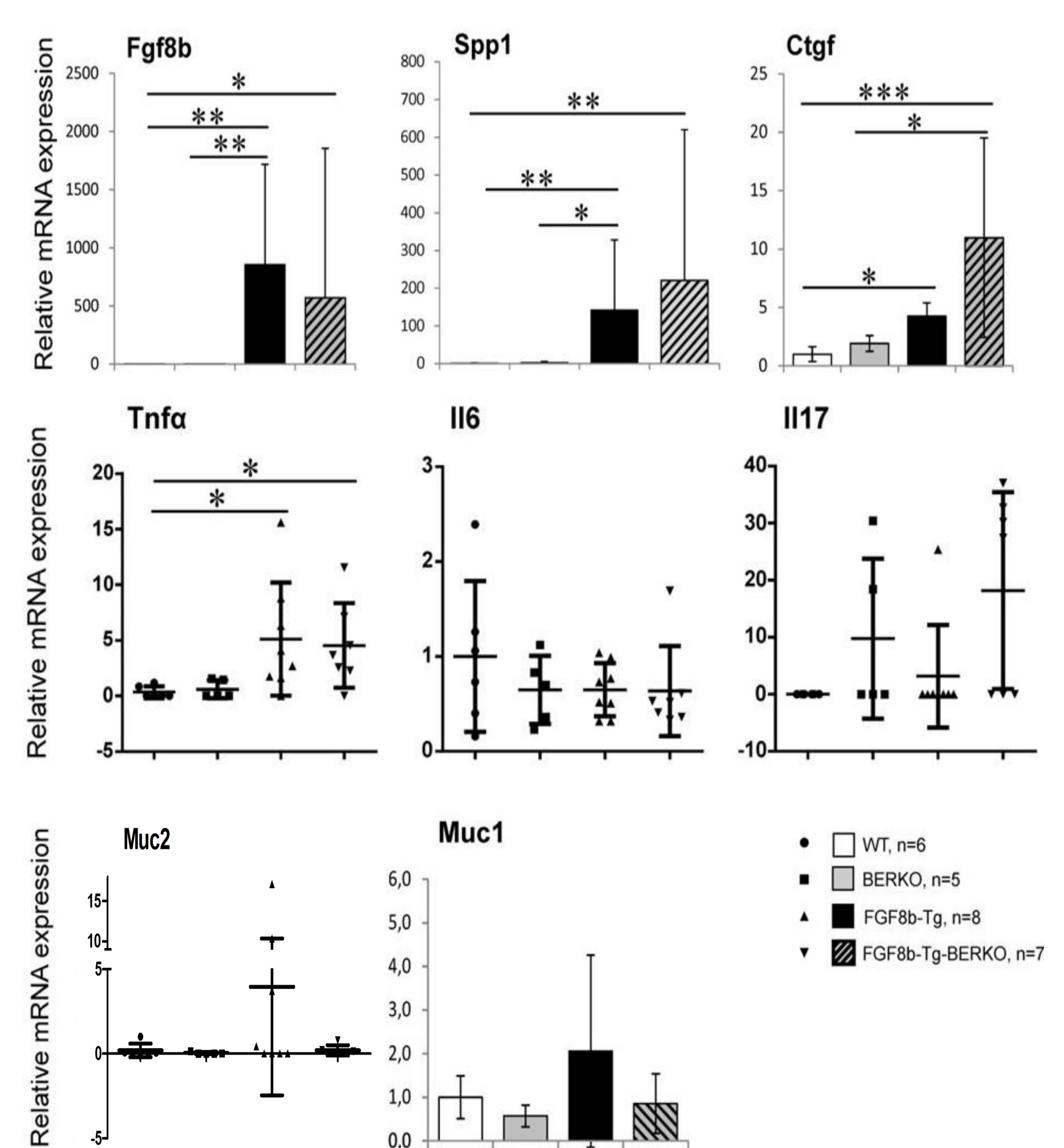
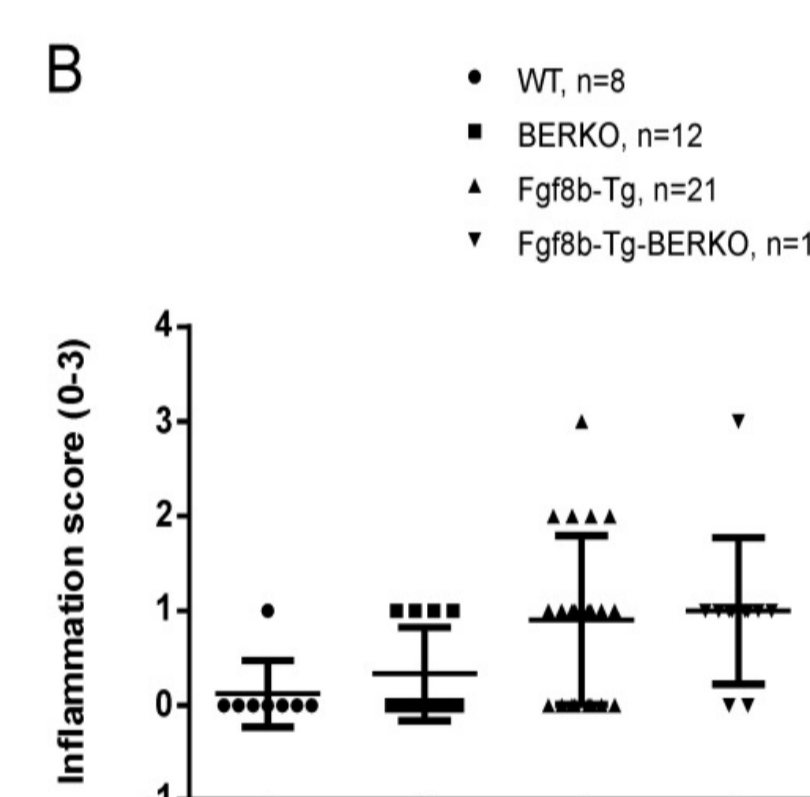
## Results



**Figure 1.** *Fgf8b*-Tg-BERKO mice (FVB/N strain) were generated by breeding female *Fgf8b*-Tg mice (*Fgf8b*<sup>+/-</sup>) with male BERKO (*Esr2*<sup>-/-</sup>) mice, which in the F1 generation gained mice heterozygotes for *Esr2* knockout (*Esr2*<sup>+/-</sup>) of which half were *Fgf8b*-Tg-positive (*Fgf8b*<sup>+/-</sup>) and half negative (*Fgf8b*<sup>-/-</sup>). Next, *Fgf8b*<sup>+/-</sup> *Esr2*<sup>+/-</sup> female mice were bred with *Fgf8b*<sup>-/-</sup> *Esr2*<sup>-/-</sup> and *Fgf8b*<sup>-/-</sup> *Esr2*<sup>+/-</sup> male mice to obtain F2 offspring with *Fgf8b*<sup>+/-</sup>, *Esr2*<sup>-/-</sup> (*Fgf8b*-Tg-BERKO); *Fgf8b*<sup>+/-</sup>, *Esr2*<sup>+/-</sup> (*Fgf8b*-Tg); *Fgf8b*<sup>-/-</sup>, *Esr2*<sup>-/-</sup> (BERKO); *Fgf8b*<sup>-/-</sup>, *Esr2*<sup>+/-</sup> (WT); *Fgf8b*<sup>+/-</sup>, *Esr2*<sup>+/-</sup> and *Fgf8b*<sup>-/-</sup>, *Esr2*<sup>+/-</sup> genotypes.



**Figure 2.** A) Frequency of histological changes in the prostates of 10-14-month-old WT, BERKO, *FGF8b*-Tg and *FGF8b*-Tg-BERKO mice. B) Inflammation score in the mouse prostates evaluated in the scale from 0 to 3. Mean value and standard deviation are shown. C) Normal histology of a 12.5-month-old WT mouse VP. D) Epithelial hypercellularity and inflammation in a 14-month-old BERKO mouse VP. E) mPIN stromal hypercellularity and inflammation in the VP of a 12-month-old *FGF8b*-Tg mouse. F) Mucinous metaplasia in the VP of a 12.5-month-old mouse. G-H) PAS stain in the VP of WT, BERKO, *FGF8b*-Tg and *FGF8b*-Tg-BERKO mice.



**Figure 3** Expression of mRNAs for selected cytokines and prostate cancer promoting genes studied by qRT-PCR. Beta-actin was used as a reference gene for data normalization and the relative values were counted using WT average as a reference artificially set to 1. In case of *Tnfa* qRT-PCR, the average of all the CT value data was artificially set to 1, because no signal could be detected in any of the WT prostates in this qRT-PCR. Mean values and standard deviations are shown. Differences between groups were tested by one-way ANOVA corrected with Tukey's multiple comparison test or by Kruskal-Wallis test corrected with Dunn's multiple comparison test. \*  $p < 0,05$ , \*\*  $p < 0,01$ , \*\*\*  $p < 0,001$ .

## Conclusion

- Prostates of one-year-old *Fgf8b*-Tg mice contained similar changes as previously reported, including stromal aberrations, mPIN lesions, inflammation and, in some cases, cancer.
- The prostates of one-year-old BERKO mice contained mild epithelial hypercellularity and inflammation, but no neoplastic changes (Fig 2).
- Prostate phenotype of *Fgf8b*-Tg-BERKO mice was mostly similar to that of the *Fgf8b*-Tg mice. However, mucinous metaplasia was statistically significantly ( $p = 0.013$ ) more frequent in the prostates of *Fgf8b*-Tg-BERKO mice than in the *Fgf8b*-Tg mice (Fig 2). However, gene analysis by qRT-PCR indicated that in *Fgf8b*-Tg mice, both *Muc1* and *Muc2* has higher expression compared to *Fgf8b*-Tg-BERKO mice (Fig 3).
- Inflammation and stromal and epithelial hypercellularity were more frequent in the prostate of *Fgf8b*-Tg-BERKO mice than in the prostate of *Fgf8b*-Tg mice (Fig 2). Although there was no statistically significant difference between these two groups, *Fgf8b*-Tg-BERKO mice showed tendency to higher *Il17* levels compared to other groups (Fig 3).
- The qRT-PCR results showed that the expression of mRNA for *Fgf8b* and the genes previously found to be upregulated in the prostates of *Fgf8b*-Tg mice, osteopontin (*Spp1*) and connective tissue growth factor (*Ctgf*) were also upregulated in *Fgf8b*-Tg-BERKO prostates (Fig 3).
- All in all, our results suggest that *Esr2* may have a role in the differentiation of prostatic epithelium and in protection from inflammation but they do not support the idea of a tumor-suppressive role for *Esr2*.